REVIEW



A review of the ecology, ecophysiology and biodiversity of microalgae in Arctic soil crusts

Ekaterina Pushkareva¹ · Jeffrey R. Johansen³ · Josef Elster^{1,2}

Received: 7 July 2015/Revised: 1 February 2016/Accepted: 3 February 2016 © Springer-Verlag Berlin Heidelberg 2016

Abstract Biological soil crusts have been extensively studied in arid lands of temperate regions, particularly semi-arid steppes and warm deserts. Arctic soil crusts have received some attention, but they are far less studied than their temperate counterparts. While the tundra zone of Arctic regions has an abundant cover of lichens, mosses and low-growing vascular plants, the High Arctic semi-arid and arid deserts have a much reduced but still very significant cover of biological soil crust dominated by microalgae. This review discusses what is known about Arctic soil crusts with the intention of stimulating study of this sensitive ecosystem. Arctic soil crusts are considered to be one of the most extreme habitat types on earth. Low temperatures and lack of water associated with a wide spectrum of disturbances have a dramatic effect on chemical and physical soil ecological properties (salinity, pH, conductivity and gas content). Microalgae are the keystone microbial species in polar crusts, being significant primary producers, fixing atmospheric nitrogen and secreting

☑ Josef Elster jelster@prf.jcu.cz

> Ekaterina Pushkareva puekse@gmail.com

Jeffrey R. Johansen johansen@jcu.edu

- ¹ Centre for Polar Ecology, Faculty of Science, University of South Bohemia, Branišovská 31, 370 05 České Budějovice, Czech Republic
- ² Centre for Phycology, Institute of Botany, Academy of Sciences of the Czech Republic, Dukelská 135, 379 82 Třeboň, Czech Republic
- ³ Department of Biology, John Carroll University, 1 John Carroll Blvd., University Heights, OH 44118, USA

polysaccharides that bind soil aggregates together, thereby reducing erosion and water runoff. The biological diversity of soil crust microalgae in the Arctic is high. Soil crusts of the Arctic semi-arid and arid deserts provide a special opportunity to study the environmental factors controlling the diversity, distribution and abundance of the microalgae in the absence of anthropogenic disturbance. However, anthropogenic disturbances and climate change are occurring in the Arctic, and even more transformations are expected in the near future. Therefore, the ecological study of Arctic ecosystems, including biological soil crusts, is a matter of urgency.

Keywords Soil crust · Arctic · Cyanobacteria and eukaryotic microalgae

Introduction

Biological soil crusts of arid and semi-arid regions have been studied over the past 35 years. Deserts and semi-arid steppes of temperate regions have received the most attention, where soil crusts have been found to have several key environmental roles, including soil stabilization and the subsequent protection from erosion by wind and water, nitrogen fixation, contribution to soil organic matter, influence on hydrology and infiltration of rain water, increasing soil temperature and affecting the mineral nutrient supply to vascular plants. Extensive work has also been conducted on the disturbance ecology of crusts, including both the impact of various types of disturbance and the recovery or succession following the cessation of the disturbance. Researchers have also been interested in the diversity of all major crust constituents (lichens, mosses, cyanobacteria, eukaryotic microalgae, fungi and heterotrophic prokaryotes). The literature dealing with crusts was reviewed in detail by Belnap and Lange (2001).

Our understanding of the biological soil crusts of polar desert and polar semi-desert regions is far less expansive. Green and Broady (2001) provided a review of the biological soil crusts of Antarctica, but at the time of this review much less was known about the ecosystem function of polar crusts. There has been considerable progress in expanding our understanding of polar biological soil crusts since these seminal reviews, for example, the recent review of soil crusts in Antarctica (Büdel and Colesie 2014). Recent reviews in Arctic terrestrial ecosystems are limited to Stewart et al. (2014) where the authors focused on atmospheric nitrogen exchanges. In addition, the recent article (Pointing et al. 2015) provides information about the biogeography of photoautotrophs in the Arctic and Antarctica.

The purpose of this article is to provide a current review of the biological soil crusts of Arctic semi-arid and arid deserts with a focus on the microalgal community. Here, we use the term "microalgae" to combine both prokaryotic cyanobacteria and eukaryotic microalgae.

Extent and components of Arctic soil crusts

Soil crusts of the Northern Hemisphere are mainly represented in the High Arctic (Fig. 1) and cover the territories of the European, Russian and Canadian Arctic as well as Greenland.

As shown in Fig. 1, the Arctic desert and semi-desert biome covers large parts of the High Arctic. However, in respect to area, the Canadian Arctic Archipelago and northeast part of the America contain most of this biome. The Canadian Arctic archipelago and neighboring coastal areas of Greenland constitute an immense geographical region dominated by polar deserts and semi-deserts (Bliss et al. 1984). The area differs in altitude, substratum and other ecological characteristics. In the spring, polar desert soils are supplied by a surge of meltwater, usually followed by longer periods of drought during summer (Elster 2002; Elster and Benson 2004). Thus, the microalgal communities are subject to seasonal extremes of inundation and desiccation, hypo- and hypersalinity, seasonal and diurnal temperature fluctuations, frequent freeze-thaw cycles and, ultimately, deep-freeze temperatures up to -34 °C in winter (Láska et al. 2012).

Even though polar desert occupies 15–20 % of the surface area of the Arctic, only a limited number of studies deal with the Arctic biological soil crust ecosystem (Liengen 1999; Elster et al. 1999; Kaštovská et al. 2005, 2007; Patova and Beljakova 2006; Breen and Lèvesque 2006, 2008; Yoshitake et al. 2007, 2010, 2014; Andreyeva

2009; Pushkareva and Elster 2013; Steven et al. 2013; Inoue et al. 2014; Pushkareva et al. 2015; Shi et al. 2015). Most of the studies describing microalgal communities in Arctic soil crusts are based on morphology (Elster et al. 1999; Kaštovská et al. 2005, 2007; Patova and Beljakova 2006; Andreyeva 2009; Pushkareva and Elster 2013). A few studies have applied modern molecular methods for microalgal identification, but most of these studies present data for the whole microbial community without a detailed assessment of microalgae (Schutte et al. 2010; Knelman et al. 2012; Steven et al. 2013; Pushkareva et al. 2015). In Table 1, we summarize the microalgal species that have been found in Arctic soil crusts. Greenland is excluded because of the lack of information on crusts of this large and important island.

The dominant components in the soil cyanobacterial community are filamentous forms (Table 1). Representatives from the orders Chroococcales, Pseudanabaenales, Oscillatoriales and Nostocales are the most species-diverse groups found in Arctic soil crusts (Kaštovská et al. 2005, 2007; Patova and Beljakova 2006; Pushkareva and Elster 2013; Pushkareva et al. 2015). Nostoc spp. are widely dispersed in all types of soil crusts and primarily inhabit the soil surface (Yeager et al. 2004; Řeháková et al. 2011; Hu et al. 2012; Bastida et al. 2014). Arctic habitats with high humidity, such as areas around streams, pools, lakes, waterfalls and snow fields, have soils frequently covered by a thick layer of *Nostoc* spp. (Pushkareva and Elster 2013). A high abundance of Nostoc spp. might also be explained by decreased nitrogen availability in the soil crusts (Zielke et al. 2005; Stewart et al. 2012). Notably absent from Arctic soils are the genera Oculatella and Hassallia, which are common in desert soil crusts from temperate and tropical regions (Flechtner et al. 2008; Patzelt et al. 2014; Osorio-Santos et al. 2014).

The extreme conditions of polar regions strongly affect the diversity of eukaryotic microalgae. Coccoid green algae (Chlorophyta) are the most abundant eukaryotic group in Arctic soil crusts (Lange 2001; Andreyeva 2009; Pushkareva and Elster 2013; see Table 1) represented by Chlorophyceae and Trebouxiophyceae (Elster et al. 1999; Andreyeva 2009). Some soils are wet and acidic and support sacoderm desmids as well (see Table 1, Zygnematophyceae). In addition to the coccoid taxa, filamentous taxa can be important as well (e.g., *Klebsormidium, Xanthonema* and *Tribonema*; see Elster 2002). Representatives of the stramenopile (heterokont) class Xanthophyceae are also abundant in polar regions (Table 1).

In the Russian Arctic the soil crust ecosystem is well described in tundra zones. However, accessibility to this literature is difficult because most of the papers are in Russian language and consequently the findings of Russian

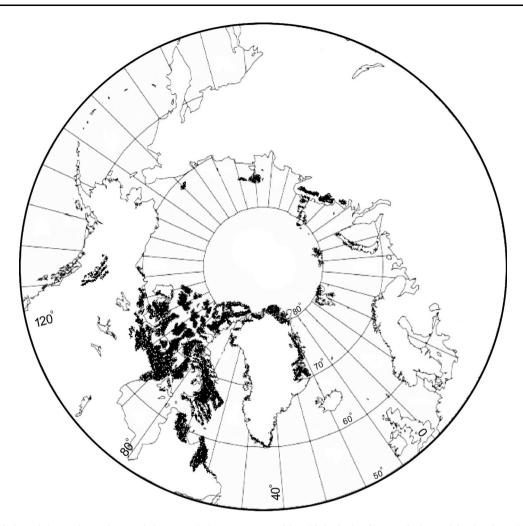


Fig. 1 Map of the High Arctic. Polar semi-desert and desert are colored *black* and represent areas where biological soil crust development is most widespread. In particular geographical areas more than 65° N, high elevation (more than 400-1000 m a.s.l.), lack of precipitation (less than 250 mm), mean air temperature lower than -10 to -15 °C, soil temperature regime (hypergolic) and continentality (eucontinental, hypercontinental), the polar desert and semi-desert with biological soil prevail (see more in the Soil Atlas of the Northern Circumpolar Region, Jones et al. 2009). The highest area

scientists are poorly known. The most important representatives of mosses in the Russian Arctic are Rhacomitrium lanuginosum, Ditrichum flexicaule and Dicranoweisia crispula (Matveeva 1979; Afonina and Matveyeva 2003). The lichen flora is dominated by the genus Cetraria (C. delisei, C. islandica var. polaris, C. laevigata, C. elenkinii) (Matveeva 1979; Zhurbenko and Matveyeva 2006). To the best of our knowledge, cyanobacterial species composition of soil crusts in the Russian Arctic was described only on Bolshevik Island, Severnaya Zemlya Archipilago (Patova and Beljakova 2006; see Table 1).

Andreyeva (2009) identified nine species of Chlorophyta from Alexandra Land (Franz Josef Land) and 22

with a biological soil crust is located in the Canadian High Arctic (southern part of Nunavut and Canadian Arctic Archipelago). A lesser extent of soil crust is also located in the northeast sea shore of Greenland, northern part of Svalbard, Franz Josef Land, Severnaya Zemlya Archipelago, and higher altitude in Taymyr Peninsula and New Siberian Islands. The southern limit is located in the southeast part of North America (Newfoundland and Labrador, Baffin Island, north part of Hudson Bay)

species from Bolshevik Island (Table 1). Common genera for both regions are *Chlamydopodium* sp. and *Chlorococcum* sp.

Ecological factors affecting the polar soil crust ecosystem

Soil crust communities can occur in almost all types of soil, and many biotic and abiotic parameters influence their development (soil geological properties, climate, presence of vascular plants, and animal and human intervention) (Langhans et al. 2009; Fischer and Subbotina 2014; Huang et al. 2014; Pushkareva et al. 2015).

	Russian Arctic (Bolshevik Island, Alexandra Land) ^{a,b}	European Arctic (Svalbard) ^{a,c,d,e}	Canadian Arctic (Ellesmere Island, Ellef Ringens Island) ^{a,f}
Cyanobacteria			
Chroococcales			
Aphanocapsa	+		
Aphanothece	+		
Chlorogloea		+	
Chroococcus	+	+	+
Cyanosarcina			+
Gloeocapsa	+	+	+
Gloeocapsopsis	+		
Gloeothece	+		
Rhabdoderma	+		
Synechococcus			+
Synechocystis	+		
Pseudanabaenales			
Geitlerinema		+	
Komvophoron		+	
Leptolyngbya	+	+	+
Phormidesmis		+	
Pseudanabaena		+	
Schizothrix			+
Oscillatoriales			
Microcoleus		+	+
Oscillatoria	+	I	
Phormidium	+	+	+
Symploca	+	I	I
Symplocastrum	+		
Nostocales	I		
Anabaena	+	+	
Calothrix	+	+	
Cylindrospermum		т	
Dichothrix	+ +		
Nodularia	+		
		+	
Nostoc	+	+	+
Scytonema	+	+	
Stigonema	+		
Tolypothrix	+	+	
Trichormus		+	
Stramenopila			
Xanthophyceae			
Botrydiopsis			+
Heterococcus			+
Monodopsis			+
Xanthonema			+
Chlorophyta			
Chlorophyceae			
Actinochloris			+
Ascochloris			+
Asterococcus			+

Table 1 Microalgal genera reported in biological soil crusts, based on seven studies of these communities in the Arctic

Table 1 continued

	Russian Arctic (Bolshevik Island, Alexandra Land) ^{a,b}	European Arctic (Svalbard) ^{a,c,d,e}	Canadian Arctic (Ellesmere Island Ellef Ringens Island) ^{a,f}
Bracteacoccus	+	+	+
Borodinellopsis			+
Chlamydocapsa	+		+
Chlamydopodium	+		+
Chlamydomonas			+
Chlorolobion			+
Chlorococcum	+	+	+
Chloromonas			+
Chlorosarcina			+
Chlorosarcinopsis		+	+
Chlorosphaeropsis			+
Coccobotrys			+
Coleochlamys		+	
Deasonia			+
Dictyochloris			+
Dictyococcus			+
Diplosphaera			+
Ettlia			+
Gloeococcus		+	+
Gloeocystis		+	+
Halochlorella	+	+	+
Hormotila		+	1
Hormotilopsis		I	+
Macrochloris	+		+
Monoraphidium			+
Mychonastes	+	+	+
Nautococcus		I	+
Neochloris			+
Neochlorosarcina		+	+
Neospongiococcum		I	+
Palmellopsis	+		+
Planktosphaeria	Ŧ		+
Pleurastrum			
Radiosphaera			+
	+		+
Scotiellopsis Spongiochloris	+		+
	+		+
<i>Tetracystis</i>	+	+	+
Frebouxiophyceae			
Chlorella	+	+	+
Choricystis		+	
Coccomyxa	+	+	
Dictyochloropsis			+
Dictyosphaerium			+
Elliptochloris			+
Keratococcus	+		+
Leptosira			+
Muriella	+	+	+
Muriellopsis	+		+

Table 1 continued

	Russian Arctic (Bolshevik Island, Alexandra Land) ^{a,b}	European Arctic (Svalbard) ^{a,c,d,e}	Canadian Arctic (Ellesmere Island, Ellef Ringens Island) ^{a,f}
Myrmecia	+	+	+
Parietochloris	+		
Pseudococcomyxa	+	+	+
Schizochlamydella			+
Stichococcus		+	+
Trebouxia		+	+
Streptophyta			
Klebsormidiophyceae			
Klebsormidium		+	+
Zygnemophyceae			
Actinotaenium			+
Cosmarium			+
Cylindrocystis		+	+
Mesotaenium			+

Diatoms are not reported, but likely they could be detected with proper methodology

^a Andreyeva (2009), ^b Patova and Beljakova (2006), ^c Kaštovská et al. (2005, 2007), ^d Pushkareva and Elster (2013), ^e Pushkareva et al. (2015), ^f Elster et al. (1999)

Excellent examples of Arctic semi-desert soil crust can be found in the northern part of Petunia Bay, Billefjorden, (78°39'22"-78°44'36"N Central Svalbard latitude, 16°22'12"-16°49'27"E longitude) (Fig. 2a, b). The highest ground cover is produced by Dryas octopetala and the lowest by the Papaver dahlianum (Prach et al. 2012). In semi-desert crusts of this area the soil surface can be consolidated by microalgae alone (Fig. 2d, f), a mixture of lichens and cyanobacteria (Fig. 2c, e), or in especially well-established crusts by a rich community of micro- or even macro-lichens (Fig. 2g, h). Such types of crusts are typical in many High Arctic areas (Belnap et al. 2001; Stewart et al. 2011b; Pushkareva et al. 2015).

In more humid habitats, the freeze-thaw cycles, occurring in soils, produce pinnacled and rolling crusts where mosses dominate (Belnap and Lange 2001; Belnap 2008). Water saturation occurs only for a short period of time during snow melt in dry, unstable soils, most frequently in steep slopes at higher elevation. Here, for most of the summer season, water is available only as water vapor from more frequent cloud occurrence. The crusts in these unstable dry soils are dominated by free-living microalgae, which usually become lichenized in less disturbed, more stable habitats (Colesie et al. 2014a). The biological crusts in temperate deserts decrease water evaporation from soil (Xiao et al. 2010; Lichner et al. 2013). This may be true for polar crusts as well, particularly those that are light-colored and do not attain elevated temperatures because of higher albedo (Fig. 3). In polar regions, the melting of snow and ice during the spring and summer periods increases the availability of liquid water (Láska et al. 2011); thus, the highest microalgal biomass presence in the Arctic is usually at the time of summer snowmelt (Elster et al. 1999).

Svalbard is a representative example of the yearly course of soil surface temperature and volumetric water content at a depth of about 2–3 cm below the surface of the soil (Fig. 3). In the Arctic semi-desert (northern part of Petunia Bay, Billefjorden, Central Svalbard) most of the year is both too cold and too dry for the growth of microalgae (October through June, Fig. 3), but both the temperature and moisture at the site shown are quite amenable to the growth of microalgae, lichens and mosses from July through September, with moisture likely being the more limiting factor (Fig. 3).

Soil texture greatly influences biological crust communities in polar regions (Colacevich et al. 2009). In contrast to deep soil, the soil crust has a greater proportion of silt and clay on or just below the surface (Breen and Lèvesque 2008). Thin clay particles adhere to the mucilaginous sheath of microalgae in the soil crust. These particles have a negative charge, which allows them to bind plant micronutrients, which have a positive charge. This process increases soil fertility (Belnap and Lange 2001; Breen and Lèvesque 2008). A more stable and softer soil texture such as gypsum and silty loams increases the diversity of microalgae, lichens and mosses (Belnap et al. 2001). The presence of unstable and coarse soils negatively influences the abundance and diversity of the structural organisms of soil crusts in the Arctic (Kaštovská et al. 2005). However, filamentous cyanobacteria can be quite abundant in the

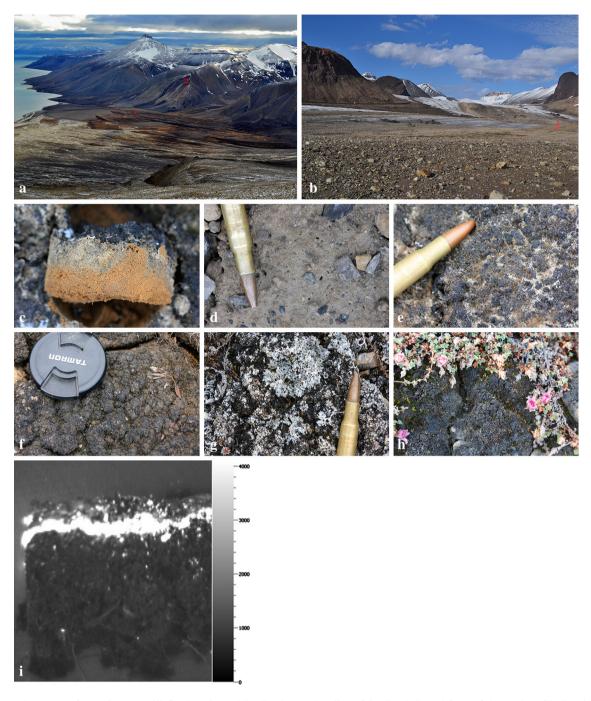


Fig. 2 Northern part of Petunia Bay, Billefjorden, Central Svalbard, Arctic semi-desert soil crust (a). Close-up images (d, e, f, g, h) show development of a soil crust community from quite young bare habitat in front of the Horbye Glacier (b, d) up to more developed (e, f) and partially covered by vascular plants and lichenized soil crust types (g, h). Arrows in images a and b show the top of Mummien Peak (1), a raised marine terrace that is characteristic of the early Holocene

coarse type of soil crust (Pushkareva et al. 2015). Welldeveloped soil crusts with a high density of soil crust organisms, mainly microalgae, lichens and mosses, are usually dark in color (Yoshitake et al. 2007), but sometimes the surface can be paler because of certain lichen shoreline of Svalbard (2) and front of the Horbye Glacier (3). Crosssections of a soil crust and its surface (c, i) show algal presence and photosynthetic activity in surface layers. An image (i) taken with the FluorCam 700MF fluorescence imaging camera (Photon Systems Instruments, Czech Republic) has a distinct whitish layer demonstrating the photosynthetically active area of soil crust. The *vertical scale* demonstrates relative units of imaging fluorometry

species, for example, *Ochrolechia frigida* (Inoue et al. 2014). In dark-colored well-developed biological crusts, there are more available mineral nutrients (such as phosphorus and/or nitrogen) and organic carbon (Chae et al. 2016; Pushkareva et al. 2015). In contrast, a poorly

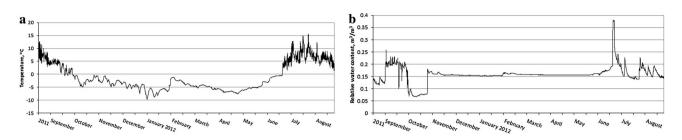


Fig. 3 Soil temperature (a) and volumetric water content (b) at a depth of 2 cm below the surface from the Herbye glacier foreland (northern part of Petunia Bay, Billefjorden, Central Svalbard) in the period from August 2011 to August 2012

developed light-colored soil surface is associated with a low biomass of major soil crust organisms followed by lower photosynthetic activity and lower pigment content (Chae et al. 2016; Pushkareva et al. 2015).

Dark-colored soil crusts better absorb long-wave radiation, which may increase the temperature of the soil surface (Stradling et al. 2002). Dark soils usually retain warmth for a longer time than light-colored soils and can positively influence the growth of biological soil crusts and vascular plants. Nevertheless, UV radiation can negatively affect Arctic soil crust organisms (Solheim et al. 2006). Microalgae can enter into a resting stage under such stressful conditions (Chen et al. 2003; Tashyreva and Elster 2012, 2015; Pichrtová et al. 2014). Motile cyanobacteria, such as Microcoleus vaginatus, M. steenstrupii and other filamentous cyanobacteria, typically occupy a position from 1 to 5 mm below the soil surface and thus can achieve a balance between receiving sufficient light for photosynthesis and avoiding harmful UV radiation and photooxidation (Castenholz and Garcia-Pichel 2000; Norris and Castenholz 2006). Non-motile cyanobacteria, such as the heterocytous genera Scytonema, produce dark yellowish pigmentation in their sheaths, which acts as a UV screen and darkens the soil surface associated with biological soil crusts (George et al. 2001).

Soil pH is a very important factor for the growth and diversity of microalgae. Soil crusts in temperate and tropical regions with a low pH (<7.0) are usually dominated by green algae (Johansen et al. 1993; Hastings et al. 2014), while the pH optimum for cyanobacteria is between 7.4 and 8.0 (Burja et al. 2002). Lower pH was found to promote higher microalgal abundance in Arctic soil crusts (Kaštovská et al. 2007). However, an increase of pH results in dominance of filamentous cyanobacteria (Pushkareva et al. 2015).

Organisms living on the soil surface are more easily exposed to and affected by wind compared to organisms inhabiting other parts of the ecosystem (Jia et al. 2012). Because of the sparse vegetation in polar regions, the soil crust organisms lack protection from the wind. Soil movements generated by freeze-thaw processes limit development of vascular plants, but likely are less limiting to microbial components of biological soil crusts (Jia et al. 2012). However, the responses of polar desert and/or semidesert biological soil crust to wind are still largely unexplored.

Adaptation of soil crust microalgae to polar conditions

In polar soil crust ecosystems, microalgae have developed a wide range of adaptive strategies that allow them to avoid, or at least minimize, the injurious effects of extreme and fluctuating environmental conditions (Elster 2002). Soil microalgae are poikilohydric microorganisms (having no mechanism to prevent desiccation), but are extremely tolerant to drought and are able to quickly return to an active state upon rehydration. They can survive long periods of desiccation and strong fluctuations in temperature (Moquin et al. 2012; Pichrtová et al. 2014; Tashyreva and Elster 2015) and have very different life strategies with respect to their susceptibility to low temperatures and freezing. This could be because cyanobacteria and several microalgae (mainly terrestrial Chlorophyceae) do not contain vacuoles, which in many eukaryotic microalgae and plants are the cellular component responsible for water control. For example, in the marine polar ecosystem vacuoles have been recorded in microalgae quite frequently (Kirst 1990; Kirst and Wiencke 1995), while there is scarce information about the presence of vacuoles in polar soil microalgae. The external environment directly manages the metabolic activity of poikilohydric organisms by affecting the presence or absence of water in either liquid or vapor form. Cyanobacteria are particularly well adapted to the severe and changeable conditions involving cycles of desiccation, rehydration, low to high salinity and freezethaw episodes. High light intensity during freezing and desiccation has been shown to negatively influence the survival of Leptolyngbya spp. (George et al. 2001). In contrast, Scytonema spp. usually live on the surface of biological soil crusts, requiring the production of the UV-

screen sheath pigment scytonemin (Quesada et al. 1999). However, the biomass of *Scytonema* sp. can decrease where mosses occur and compete for light (Lan et al. 2012).

Microalgae are the most important photosynthetic organisms of soil crusts in the polar regions (Elster 2002). However, they often have a low concentration of photosynthetic pigments per unit area (Elster et al. 1999) as a consequence of the harsh climatic and environmental conditions (Colacevich et al. 2009). Figure 2c, i shows photosynthetic organisms (mainly microalgae) living on the soil surface or slightly below it. Because of structural and functional differences in photosynthetic organelles, eukaryotic microalgae have higher rates of photosynthesis and lower resistance to freeze-thaw cycles than cyanobacteria (Šabacká and Elster 2006; Pichrtová et al. 2014). A good example of structural changes in cell morphology that provide resistance to freeze-thaw injuries was documented in a study of the annual development of matforming conjugating green algae, Zygnema sp. (Pichrtová et al., in press), showing that in summer Zygnema sp. cells contain stellate chloroplasts and large hyaline vacuoles. As summer progressed, Zygnema sp. produced pre-akinetes, overwintering cells, with high lipid content and reduced chloroplast lobes. Such cells have been found to be able to survive winter conditions and other extremes. However, features responsible for resistance against low temperature, cryoinjuries, desiccation and salinity stress are taxonomically specific at the species and ecoform levels (Elster and Benson 2004). Cyanobacteria, in contrast to eukaryotic microalgae, have lower rates of photosynthesis, but their biomass production is higher than their subsequent decomposition. Accumulation of cyanobacterial biomass in Arctic soil crust habitats is, in addition to the low rate of decomposition, influenced by low grazing pressure by invertebrates. However, these processes are still poorly understood, and accumulation of cyanobacterial biomass in particular polar habitats needs further research. For example, soil crusts dominated by Nostoc spp. rapidly rebuild (during several hours or days) after freezing and desiccation (Hawes et al. 1992). Extracellular polysaccharides in the sheath and colonial investments of cyanobacteria often persist in the environment and can serve to aggregate soil particles and stabilize the soil surface even after the cellular components of the soil have died (Elster and Benson 2004; Tashyreva and Elster 2015).

Soil crust microalgae, due to a diverse range of ecological and physiological life strategies, manifest an ability to tolerate stressful conditions. There are three main strategies for survival in polar soil crust habitats: avoidance of stress, protection from stress and forming partnerships with other organisms. Water availability and state (liquidice-vapor) connected with all types of mechanical disturbances or instability are decisive factors that determine whether avoidance, protection or formation of a partnership prevails. In conditions where there is a more regular occurrence of water in the liquid form, avoidance or protection is more common. In contrast, protection or forming of partnerships is quite common in habitats where water in liquid form occurs only for a limited time or where water is primarily available only in vapor form (Elster 2002).

Some soil crust microalgae are motile in vegetative and/ or reproductive cell stages. Mobility can facilitate avoidance of the most stressful conditions and propel the organism to a more favorable environment. Motile microalgae can also react to a wide spectrum of environmental conditions (soil properties, temperature, moisture). The mucilage protects them against fluctuations in water status by inhibiting water loss from cells. When soil is wet, the mucilage of cyanobacteria swells and trichomes migrate out of their sheaths (Hu et al. 2012). After each migration, new sheath material is formed, thus extending the filament length. Repeated swelling leaves a complex network of empty sheath material, which maintains the soil structure after the organisms have become dehydrated and decreased in size.

Other strategies for survival in severe conditions are the development of resting (dormant) vegetative stages as well as reproductive stages that affect an organism's ability to adapt to seasonal environmental fluctuations (Tashyreva and Elster 2012, 2015). We have already mentioned production of pre-akinetes in Arctic populations of Zvgnema sp. (Pichrtová et al. 2014). In contrast, Phormidium sp. (Oscillatoriales, Cyanobacteria) forms perennial populations, with a high proportion of cells able to survive winter without specialized cells being produced (Tashyreva and Elster 2016). This is a very important adaptive feature for living in polar conditions. Physiological changes preceding dormancy include accumulating high concentrations of soluble carbohydrates that substitute for water molecules during dehydration (Elster and Benson 2004). These intracellular carbohydrates stabilize the structure and function of macromolecules, membranes and cellular organization. The presence of protective sugars in cells enables vitrification of cytoplasm on drying and supports the formation of a high-viscosity, metastable glassy state. This preserves cell viability during dry frozen storage by immobilizing cellular constituents and suppressing deleterious chemical or biochemical reactions that threaten survival (Sun and Leopold 1994).

An important feature of soil crust communities in polar regions is the production of life-form associations such as mutualistic symbiosis, which offer protection against unstable extreme conditions on the soil surface layer. They are traditionally defined as the living together of two or more unlike organisms to the benefit of all partner organisms. The life associations that commonly occur in the polar terrestrial environment, with the exception of physical protection, also have physiological and metabolic advantages.

The most frequent and most ecologically important association in the polar terrestrial environment is the association of microalgae with fungi in lichens (Bjerke 2011; Inoue et al. 2011, 2014). Fungal hyphae are frequent components of the cryptogamic crust. Through their filamentous hyphae, fungi in crusts contribute to soil stability by aggregating soil particles (Abed et al. 2012). This symbiosis helps to protect them from low temperatures and water fluctuations (Stewart et al. 2011b). Lichens that use green algae as photobionts are usually dominant in polar soil crusts. Lichens that have the green alga Trebouxia sp. as the photobiont are known to be extremely cold-resistant in the dry as well as hydrated states (Leal 2000), and several lichen species are known to be able to attain significant net photosynthetic rates at subzero temperatures.

Another important and common example of association in Arctic soil crusts is epiphytic occurrence of cyanobacteria on moss surfaces (Stewart et al. 2011b). Probably, the benefit for cyanobacteria in this kind of association may be the supply of carbohydrates and protection against desiccation and UV radiation while mosses can get nitrogen fixed from the cyanobacteria (Zielke et al. 2005).

Ecological role of microalgae in soil crusts

Soil crust organisms are involved in important processes of polar soil ecosystems such as nitrogen fixation, moisture retention, stabilizing of soil and increasing of soil organic carbon (Breen and Lévesque 2008; Stewart et al. 2012, 2014; Büdel and Colesie 2014; Chae et al. 2016). Stabilization of the soil surface by soil crust organisms helps to protect the soil from erosion through both aggregation of the soil and speeding the rate of water infiltration (Colesie et al. 2014b). By increasing surface roughness, they reduce runoff and, as a consequence, increase infiltration and the amount of water stored for plant use. Cyanobacteria secrete polysaccharides that bind soil, thus influencing the soil stability, erosion, runoff and growth of soil crust components such as lichens (Colesie et al. 2014a).

A major feature of cyanobacteria is the ability of heterocystous species to fix atmospheric nitrogen (Solheim et al. 2006; Maqubela et al. 2009; Stewart et al. 2011a). They start to fix nitrogen as soon as the temperature in the thallus rises above 0 °C. In addition to accumulation of organic carbon and other elements in cells, nitrogen fixation is one of the most important ecological contributions to cyanobacteria-rich Arctic soil crusts. Nitrogen-fixing bacteria (including cyanobacteria) are a significant source of fixed nitrogen for plants and edaphic heterotrophic microorganisms. Both free-living forms and those associated with species of vascular plants have been reported as being important nitrogen sources in High Arctic locations (Zielke et al. 2005; Breen and Lèvesque 2006). In nature, nitrogen-fixing cyanobacteria are abundant in areas deficient in nitrogen.

Nitrogen fixation depends on the water availability, temperature, soil chemistry and other parameters (Zielke et al. 2005; Kvíderová et al. 2011). For example, soil crusts with a sandy cover can contain higher concentrations of nitrogen compared with non-sand-covered soil crusts (Williams and Eldridge 2011). However, the relation of the rate of nitrogen fixation and texture and the type of soil substrate has not yet been studied. Phosphate limitation can negatively affect nitrogen fixation by decreasing the rate of photosynthesis and consequently inhibiting nitrogenase by reducing the photosynthates required for the energy-intensive process of nitrogen fixation (Hartley and Schlesinger 2002; Stewart et al. 2011a).

Cyanobacteria are often responsible for the majority of carbon fixed in Arctic soils, providing fixed carbon to soil crusts, which likely subsidizes food webs there (Yoshitake et al. 2010; Darby et al. 2010). In addition, the abundance of cyanobacteria increases with crust development, resulting in a higher carbon concentration in well-developed soil crusts rather than in poorly developed ones (Kaštovská et al. 2007; Pushkareva et al. 2015).

Endemism or cosmopolitanism?

The question about endemism of polar terrestrial microorganisms is still unsettled (Lawley et al. 2004; Casamatta et al. 2005; Rybalka et al. 2009; Strunecký et al. 2012) because many isolates found in the Arctic and the Antarctic share similar morphology with microalgae from other geographic regions. Microorganisms can be transported by dispersed vectors such as atmospheric circulation, ocean currents, animals (particularly migratory birds) and humans (Lawley et al. 2004; Strunecký et al. 2010). However, recent molecular analyses have not detected totally identical genotypes between particular species of polar regions and other locations (Lawley et al. 2004; Rybalka et al. 2009; Schmidt et al. 2011; Strunecký et al. 2010, 2012). It is very possible that the unique challenges of polar ecosystems prevent establishment by most microbial introductions and that the microorganisms inhabiting these regions have become specially adapted.

The increase in scientific and tourist activities in polar regions has a potentially huge impact on the geographical distribution of microalgae genotypes. Anthropogenic activities promote species movement and their subsequent reproduction and spread at the new locations (Walther et al. 2002). Newly transported genotypes may adversely interfere with indigenous species and endanger their subsistence. Human presence in all polar regions has increased, thereby extending threats to biodiversity. However, up to now we have had very limited information about anthropogenic impacts on the geographical distribution of microalgae. There is a complete lack of data on alien microalgae dispersal and their subsequent occupation of particular localities and habitats. With the concomitant rapid climate change occurring in polar areas (particularly the Arctic and northwest Antarctic Peninsula region), the locally adapted species may lose their competitive advantage and be displaced by invasive species (Frenot et al. 2005). However, recent research on the dispersal of morphologically simple airborne eukaryotic microalgae, Klebsormidium (Streptophyta), Chlorella and Stichococcus (Chlorophyta), to polar regions showed ubiquitous distribution on a global scale (Hodač et al., in press; Ryšánek et al., in press). Almost 80 % of all Arctic Klebsormidium strains were included within the cosmopolitan superclade B sensu Rindi et al. (2011). Similarly, Hodač et al. (in press) found that psychrotolerant strains of Chlorella and Stichococcus are without exception conspecific (or closely related) with strains originating from the temperate zone. A warming climate can promote immigration of alien species into the polar regions and could cause shifts in species abundance and distribution. The combined effects of invasive species and climate change on the biodiversity of soil organisms could modify the polar ecosystem.

Conclusion

This article gives a brief overview of the microalgal ecology, physiological ecology and biodiversity of soil crust ecosystems in polar regions with a focus on Arctic soil crusts. Low temperature, low availability of liquid water and instability are characteristic for these terrestrial ecosystems. A combination of ecological properties, including geochemical and physical factors, light availability, grazing pressure by invertebrates, anthropogenic impacts and invasive species, influences various microalgal processes in soil crust ecosystems. The severe conditions influence the diversity, abundance and ecological manifestation of organisms in soil crusts. Soil microalgae have developed diverse ecological and physiological life strategies and behaviors that help them to avoid stressful conditions in the upper soil layer. Survival strategies in these microorganisms (similar to organisms living in temperate and tropical regions) have occurred because of the considerable evolutionary pressures experienced and an exceptionally long period of predictably unpredictable climatic conditions. Study of the community composition and main processes in Arctic soil crusts is necessary for better prediction of the future climate change.

Acknowledgments We are indebted to Kamil Láska, PhD (Masaryk University, Brno), and Jan Kavan, MSc (University of South Bohemia, České Budějovice), for measuring of soil microclimatic parameters. Our study was supported by grants from the Ministry of Education, Youth and Sport of the Czech Republic (LM2010009 CzechPolar, CZ.1.07/2.2.00/28.0190 and RVO67985939). Special appreciation is given to reviewer Prof. P. Broady, who helped immensely to increase the quality of the manuscript.

References

- Abed RMM, Al-Sadi AM, Al-Shehi M, Al-Hinai S, Robinson MD (2012) Diversity of free-living and lichenized fungal communities in biological soil crusts of the Sultanate of Oman and their role in improving soil properties. Soil Biol Biochem 57:695–705
- Afonina OM, Matveyeva NV (2003) Mosses of the Bolshevik Island (Severnaya zemlya Archipilago). Bot J 88(9):1 (in Russian)
- Andreyeva VM (2009) Nonmotile unicellular and colonial green algae (chlorophyta) in soils of polar deserts. Novit Syst Plant Non Vasc 43:7–15 (in Russian)
- Bastida F, Jehmlich N, Ondono S, Bergen M, Garcia C, Moreno JL (2014) Characterization of the microbial community in biological soil crusts dominated by *Fulgensia desertorum* (Tomin) Poelt and *Squamarina cartilaginea* (With.) P. James and in the underlying soil. Soil Biol Biochem 76:70–79
- Belnap J (2008) Biological Crusts. In: Lal R (ed) Encyclopedia of soil science. Taylor & Francis, New York, pp 1–4
- Belnap J, Lange OL (2001) Biological soil crusts: structure, function and management. Springer, Berlin
- Belnap J, Rosentreter R, Kaltenecker J, Williams J, Leonard S, Luehring P, Eldridge D (2001) Biological soil crust: ecology and management. Ecology and management of microbiotic soil crusts. Technical reference, Denver
- Bjerke JW (2011) Winter climate change: ice encapsulation at mild subfreezing temperatures kills freeze-tolerant lichens. Environ Exp Bot 72:404–408
- Bliss LC, Svoboda J, Bliss DI (1984) Polar deserts and their plant cover and plant production in the Canadian High Arctic. Holarct Ecol 7:304–324
- Breen K, Lèvesque E (2006) Proglacial succession of biological soil crusts and vascular plants: biotic interactions in the High Arctic. Can J Bot 84(11):1714–1731
- Breen K, Lèvesque E (2008) The influence of biological soil crusts on soil characteristics along a High Arctic glacier foreland, Nunavut, Canada. Arct Antarct Alp Res 40(2):287–297
- Büdel B, Colesie C (2014) Biological soil crusts. In: Cowan D (ed) Antarctic terrestrial microbiology: physical and biological properties of Antarctic soil habitats. Springer, Heidelberg, pp 131–161
- Burja AM, Abu-Mansour E, Banaigs B, Pyari C, Burgess JG, Wright PC (2002) Culture of marine cyanobacterium, *Lyngbya majuscula* (Oscillatoriaceae), for bioprocess intensified production of cyclic and linear lipopeptides. J Microbiol Methods 48:207–219
- Casamatta DA, Johansen JR, Vis ML, Broadwater ST (2005) Molecular and morphological characterization of ten polar and

near-polar strains within the Oscillatoriales (Cyanobacteria). J Phycol 41:421–438

- Castenholz RW, Garcia-Pichel F (2000) Cyanobacterial responses to UV-radiation. In: Whitton BA, Potts M (eds) The ecology of cyanobacteria. Kluwer, Dordrecht, pp 591–611
- Chae N, Kang H, Kim Y, Hong SG, Lee BY, Choi T (2016) CO₂ efflux from the biological soil crusts of the High Arctic in a later stage of primary succession after deglaciation, Ny-Ålesund, Svalbard, Norway. Appl Soil Ecol 98:92–102
- Chen Y-H, Miller JR, Francis JA, Russel GL, Aires F (2003) Observed and modeled relationships among Arctic climate variables. J Geophys Res 108:D24
- Colacevich A, Caruso T, Borghini F, Bargagli R (2009) Photosynthetic pigments in soils from northern Victoria Land (continental Antarctica) as proxies for soil algal community structure and function. Soil Biol Biochem 41:2105–2114
- Colesie C, Green ATG, Haferkamp I, Büdel B (2014a) Habitat stress initiates changes in composition, CO₂ gas exchange and C-allocation as life traits in biological soil crusts. ISME J 8(10):2104–2115
- Colesie C, Gommeaux M, Green ATG, Budel B (2014b) Biological soil crusts in continental Antarctica: Garwood Valley, southern Victoria Land, and Diamond Hill, Darwin Mountains region. Antarct Sci 26(2):115–123
- Darby B, Neher D, Belnap J (2010) Impact of biological soil crusts and desert plants on soil microfaunal community composition. Plant Soil 328:421–431
- Elster J (2002) Ecological classification of terrestrial algal communities of polar environment. In: Beyer L, Boelter M (eds) GeoEcology of terrestrial oases ecological studies. Springer, Berlin, pp 303–319
- Elster J, Benson E (2004) Chapter 3. Life in the polar terrestrial environment with a focus on algae and cyanobacteria. In: Fuller JB, Lane N, Benson EE (eds) Life in a frozen state Libro. CRC Press, Boca Raton, pp 111–150
- Elster J, Lukešová A, Svoboda J, Kopecký J, Kanda H (1999) Diversity and abundance of soil algae in the polar desert, Sverdrup Pass, central Ellesmere Island. Polar Rec 35(194):231–254
- Fischer T, Subbotina M (2014) Climatic and soil texture threshold values for cryptogamic cover development: a meta analysis. Biologia 69(11):1520–1530
- Flechtner VR, Johansen JR, Belnap J (2008) The biological soil crusts of the San Nicolas Island: enigmatic algae from a geographically isolated ecosystem. West N Am Nat 68(4):405–436
- Frenot Y, Chown SL, Whinam J, Selkirk PM, Convey P, Skotnicki M, Bergstrom DM (2005) Biological invasions in the Antarctic: extent, impacts and implications. Biol Rev Camb Philos Soc 80:45–72
- George AL, Murray AW, Montiel PO (2001) Tolerance of Antarctic cyanobacterial mats to enhanced UV radiation. FEMS Microbiol Ecol 37:91–101
- Green TGA, Broady PA (2001) Biological soil crusts of Antarctica. In: Belnap J, Lange OL (eds) Biological soil crusts: structure, function, and management. Springer, Heidelberg, pp 133–139
- Hartley AE, Schlesinger WH (2002) Potential environmental controls on nitrogenase activity in biological crusts of northern Chihuahuan Desert. J Arid Environ 52:293–304
- Hastings KL, Smith LE, Lindsey ML, Blotsky LC, Downing GR, Zellars DQ, Downing JK, Corena-McLeod M (2014) Effect of microalgae application on soil algal species diversity, cation exchange capacity and organic matter after herbicide treatments [version 1; referees: 1 approved, 1 not approved]. F1000Research 3:281
- Hawes I, Howard-Williams C, Vincent WF (1992) Desiccation and recovery of Antarctic cyanobacterial mats. Polar Biol 12:587–594

- Hodač L, Hallmann C, Spitzer K, Elster J, Fashauer F, Brinkmann N, Lepka D, Diwan V, Friedl T (in press) Phylogenetic analysis of polar *Chlorella* and *Stichococcus* suggests biogeography of airborne microalgae. FEMS Microbiol Ecol
- Hu C, Gao K, Whitton BA (2012) Semi-arid regions and deserts. In: Whitton BA (ed) Ecology of cyanobacteria II: their diversity in space and time. Springer, Berlin, pp 345–369
- Huang L, Zhang Z, Li X (2014) Carbon fixation and influencing factors of biological soil crusts in a revegetated area of the Tengger Desert, northern China. J Arid Land 6(6):725–734
- Inoue T, Kudoh S, Inoue M, Uchida M, Kanda H (2011) Three lecideoid lichens new to Svalbard, Norway. Polar Sci 4(4):588–592
- Inoue T, Kudoh S, Uchida M, Tanabe Y, Inoue M, Kanda H (2014) Effects of substrate differences on water availability for Arctic lichens during the snow-free summers in the High Arctic glacier foreland. Polar Sci 8:397–412
- Jia RL, Li XR, Liu LC, Gao YH, Zhang XT (2012) Differential wind tolerance of soil crust mosses explains their micro-distribution in nature. Soil Biol Biochem 45:31–39
- Johansen JR, Ashley J, Rayburn WR (1993) The effects of rangefire on soil algal crusts in semiarid shrub-steppe of the Lower Columbia Basin and their subsequent recovery. Great Basin Nat 53:73–88
- Jones A, Stolbovoy V, Tarnocai C, Broll G, Spaargaren O, Montanarella L (eds) (2009) Soil atlas of the Northern Circumpolar Region. European Commission, Office for Official Publications of the European Communities, Luxembourg
- Kaštovská K, Elster J, Stibal M, Šantrůčková H (2005) Microbial Assemblages in soil microbial succession after glacial retreat in Svalbard (High Arctic). Microb Ecol 50(3):396–407
- Kaštovská K, Stibal M, Šabacká M, Černá B, Šantrůčková H, Elster J (2007) Microbial community structure and ecology of subglacial sediments in two polythermal Svalbard glaciers characterized by epifluorescence microscopy and PLFA. Polar Biol 30:277–287
- Kirst GO (1990) Salinity tolerance of eukaryotic marine algae. Annu Rev Plant Physiol Plant Mol Biol 41:21–53
- Kirst GO, Wiencke Ch (1995) Ecophysiology of polar algae. J Phycol 51:181–199
- Knelman JE, Legg TM, O'Neill SP, Washenberger CL, Gonzalez A, Cleveland CC, Nemergut DR (2012) Bacterial community structure and function change in association with colonizer plants during early primary succession in a glacier forefield. Soil Biol Biochem 46:172–180
- Kvíderová J, Elster J, Šimek M (2011) In situ response of Nostoc commune s.l. colonies to desiccation, in Central Svalbard, Norwegian High Arctic. Fottea 11(1):87–97
- Lan S, Wu L, Zhang D, Hu C (2012) Composition of photosynthetic organisms and diurnal changes of photosynthetic efficiency in algae and moss crusts. Plant Soil 351:325–336
- Lange OL (2001) Photosynthesis of soil crust-biota as dependent on environmental factors. In: Belnap J, Lange OL (eds) Biological soil crusts: structure, function and management. Ecological studies, vol 150. Springer, Berlin, pp 217–240
- Langhans T, Storm C, Schwabe A (2009) Biological soil crusts and their microenvironment: impact on emergence, survival and establishment of seedlings. Flora 204(2):157–168
- Láska K, Barták M, Hájek J, Prošek P, Bohuslavová O (2011) Climatic and ecological characteristics of deglaciated area of James Ross Island, Antarctica, with a special respect to vegetation cover. Czech Polar Reports 1:49–62
- Láska K, Witoszova D, Prošek P (2012) Weather patterns of the coastal zone of Petuniabukta, central Spitsbergen in the period 2008–2010. Pol Polar Res 33(4):297–318
- Lawley B, Ripley S, Bridge P, Convey P (2004) Molecular analysis of geographic patterns of eukaryotic diversity in Antarctic soils. Appl Environ Microbiol 70(10):5963–5972

- Leal GD (2000) Constraints to nitrogen fixation by cryptogamic crusts in a polar desert ecosystem, Devon Island, N.W.T. Canada. Arct Antarct Alp Res 32:40–45
- Lichner L, Hallett PD, Drongová Z, Czachor H, Kováčik L, Mataix-Solera J, Homolák M (2013) Algae influence the hydrophysical parameters of a sandy soil. Catena 108:58–68
- Liengen T (1999) Environmental factors influencing the nitrogen fixation activity of free-living terrestrial cyanobacteria from a high arctic area, Spitsbergen. Can J Microbiol 45:573–581
- Maqubela MP, Mnkeni PNS, Issa OM, Pardo MT, D'Acqui LP (2009) Nostoc cyanobacterial inoculation in South African agricultural soils enhances soil structure, fertility and maize growth. Plant Soil 315:79–92
- Matveeva NV (1979) Vegetation structure of polar desert in Taimyr Peninsula (Cape Chelyuskin). In: Aleksandrova VD, Matveeva NV (eds) Arctic tundra and polar desert of Taimyr. Nauka, Leningrad, pp 5–27 (in Russian)
- Moquin SA, Garcia JR, Brantley SL, Takacs-Vesbach CD, Shepherd UL (2012) Bacterial diversity of bryophyte-dominant biological soil crusts and associated mites. J Arid Environ 87:110–117
- Norris TB, Castenholz RW (2006) Endolithic photosynthetic communities within ancient and recent travertine deposits in Yellowstone National Park. FEMS Microbiol Ecol 57:470–483
- Osorio-Santos K, Pietrasiak N, Bohunicka M, Miscoe LH, Kovacik L, Martin MP, Johansen JR (2014) Seven new species of *Oculatella* (Pseudanabaenales, Cyanobacteria): taxonomically recognizing cryptic diversification. Eur J Phycol 49(4):450–470
- Patova EN, Beljakova RN (2006) Terrestrial cyanoprokaryota of Bolshevik island (Severnaya zemlya Archipilago). Novit Syst Plant Non Vasc 40:83–91 (in Russian)
- Patzelt DJ, Hodac L, Friedl T, Pietrasiak N, Johansen J (2014) Biodiversity of soil cyanobacteria in the hyper-arid Atacama desert, Chile. J Phycol 50:698–710
- Pichrtová M, Hájek T, Elster J (2014) Osmotic stress and recovery in field populations of *Zygnema* sp. (Zygnematophyceae, Streptophyta) on Svalbard (High Arctic) subjected to natural desiccation. FEMS Microbiol Ecol 89(2):270–280
- Pichrtová M, Hájek T, Elster J (in press) Annual development of matforming conjugating green algae *Zygnema* spp. in hydroterrestrial habitats in the Arctic. Polar Biol
- Pointing SB, Büdel B, Convey P, Gillman LN, Körner C, Leuzinger S, Vincent WF (2015) Biogeography of photoautotrophs in the high polar biome. Front Plant Sci 6:692
- Prach K, Klimešová J, Košnar J, Redcenko O, Hais M (2012) Variability of contemporary vegetation around Petuniabukta, central Spitsbergen. Pol Polar Res 33(4):383–394
- Pushkareva E, Elster J (2013) Biodiversity and ecological typification of cryptogamic soil crust in the vicinity of Petunia Bay, Svalbard. Czech Polar Rep 3(1):7–18
- Pushkareva E, Pessi IS, Wilmotte A, Elster J (2015) Cyanobacterial community composition in Arctic soil crusts at different stages of development. FEMS Microbiol Ecol 91:fiv143
- Quesada A, Vincent WF, Lean DRS (1999) Community and pigment structure of Arctic cyanobacterial assemblages: the occurrence and distribution of UV-absorbing compounds. FEMS Microbiol Ecol 28:315–323
- Řeháková K, Chlumska Z, Doležal J (2011) Soil cyanobacterial and microalgal diversity in dry mountains of Ladakh, NW Himalaya, as related to site, altitude, and vegetation. Microb Ecol 62:337–346
- Rindi F, Mikhailyuk TI, Sluiman HJ, Friedl T, Lopez-Bautista JM (2011) Phylogenetic relationships in *Interfilum* and *Klebsormidium* (Klebsormidiophyceae, Streptophyta). Mol Phylogenet Evol 58:218–231
- Rybalka N, Andersen NA, Kostikov I, Mohr KI, Massalski A, Olech M, Friedl T (2009) Testing for endemism, genotypic diversity

and species concepts in Antarctic terrestrial microalgae of the Tribonemataceae (Stamenopiles, Xanthophyceae). Environ Microbiol 11(3):554–565

- Ryšánek D, Elster J, Kováčik L, Škaloud P (in press) Diversity and dispersal capacities of a terrestrial algal genus Klebsormidium (Streptophyta) in polar regions. FEMS Micrologiol Ecol
- Šabacká M, Elster J (2006) Response of cyanobacteria and algae from Antarctic wetland habitats to freezing and desiccation stress. Polar Biol 30(1):31–37
- Schmidt SK, Lynch RC, King AJ, Karki D, Robeson MS, Nagy L, Williams MW, Mitter MS, Freeman KR (2011) Phylogeography of microbial phototrophs in the dry valleys of the high Himalayas and Antarctica. Proc Biol Sci 278:702–708
- Schutte UME, Abdo Z, Foster J, Ravel J, Bunge J, Solheim B, Forney LJ (2010) Bacterial diversity in a glacier foreland of the high Arctic. Mol Ecol 19(1):54–66
- Shi Y, Grogan P, Sun H, Xiong J, Yang Y, Zhou J, Chu H (2015) Multi-scale variability analysis reveals the importance of spatial distance in shaping Arctic soil microbial functional communities. Soil Biol Biochem 86:126–134
- Solheim B, Zielke M, Bjerke JW, Rozema J (2006) Effects of enhanced UV-B radiation on nitrogen fixation in arctic ecosystems. Plant Ecol 182:109–118
- Steven B, Lionard M, Kuske CR, Vincent WF (2013) High bacterial diversity of biological soil crusts in water tracks over permafrost in the high Arctic Polar Desert. PLoS ONE 8(8):e71489
- Stewart KJ, Coxson D, Siciliano SD (2011a) Small-scale spatial patterns in N₂-fixation and nutrient availability in an arctic hummock–hollow ecosystem. Soil Biol Biochem 43:133–140
- Stewart KJ, Lamb EG, Coxson DS, Siciliano SD (2011b) Bryophytecyanobacterial associations as key factor in N₂-fixation across the Canadian Arctic. Plant Soil 344(1-2):335-346
- Stewart KJ, Brummell ME, Coxson DS, Siciliano SD (2012) How is nitrogen fixation in the high arctic linked to greenhouse gas emissions? Plant Soil. doi:10.1007/s11104-012-1282-8
- Stewart KJ, Grogan P, Coxson DS, Siciliano SD (2014) Topography as a key factor driving atmospheric nitrogen exchanges in arctic terrestrial ecosystems. Soil Biol Biochem 70:96–112
- Stradling DA, Thygerson T, Walker JA, Smith BN, Hansen LD, Criddle RS, Pendleton RL (2002) Cryptogamic crust metabolism in response to temperature, water vapor, and liquid water. Thermochim Acta 394:219–225
- Strunecký O, Elster J, Komárek J (2010) Phylogenetic relationships between geographically separate *Phormidium* cyanobacteria: is there a link between north and south polar regions? Polar Biol 33:1419–1428
- Strunecký O, Elster J, Komárek J (2012) Molecular clock evidence for survival of Antarctic cyanobacteria (Oscillatoriales, *Phormidium autumnale*) from Paleozoic times. FEMS Microbiol Ecol 82:482–490
- Sun WQ, Leopold AC (1994) Glassy state and seed storage stability: a viability equation analysis. Ann Bot 74:601–604
- Tashyreva D, Elster J (2012) Production of dormant stages and stress resistance of polar cyanobacteria. In: Hanslmeier A, Kempe S, Seckbach J (eds) Life on Earth and other planetary bodies. Springer, Dordrecht, pp 367–386
- Tashyreva D, Elster J (2015) Effect of nitrogen starvation on tolerance of Arctic *Microcoleus* strains (Cyanobacteria) to complete and incomplete desiccation. Front Microbiol 6(278): 1–11
- Tashyreva D, Elster J (2016) Annual cycles of two cyanobacterial mat communities in hydro-terrestrial habitats of the High Arctic. Microb Ecol. doi:10.1007/s00248-016-0732-x
- Walther G-R, Post E, Convey P, Menzel A, Parmesan C, Beebee TJC, Fromentin J-M, Hoegh-Guldberg O, Bairlein F (2002) Ecological responses to recent climate change. Nature 416:389–395

- Williams WJ, Eldridge DJ (2011) Deposition of sand over a cyanobacterial soil crust increases nitrogen bioavailability in a semi-arid woodland. Appl Soil Ecol 49:26–31
- Xiao B, Zhao Y-G, Shao M-A (2010) Characteristics and numeric simulation of soil evaporation in biological soil crusts. J Arid Environ 74:121–130
- Yeager CM, Kornosky JL, Housman DC, Grote EE, Belnap J, Kuske CR (2004) Diazotrophic community structure and function in two successional stages of biological soil crusts from the Colorado Plateau and Chihuahuan Desert. Appl Eviron Microbiol 70(2):973–983
- Yoshitake S, Uchida M, Koizumi H, Nakatsubo T (2007) Carbon and nitrogen limitation of soil microbial respiration in a High Arctic successional glacier foreland near Ny-Ålesund, Svalbard. Polar Res 26:22–30
- Yoshitake S, Uchida M, Koizumi H, Kanda H, Nakatsubo T (2010) Production of biological soil crust in the early stage of primary succession on a High Arctic glacier foreland. New Phytol 186:451–460
- Yoshitake S, Uchida M, Ohtsuka T, Kanda H, Koizumi H, Nakatsubo T (2014) Vegetation development and carbon storage on a glacier foreland in the High Arctic, Ny-Alesund, Svalbard. Polar Sci 5:391–397
- Zhurbenko MP, Matveyeva NV (2006) Terricolous lichens of the Bolshevik island (Severnaya zemlya Archipilago). Botany Journal 91(10):1457–1484 (in Russian)
- Zielke M, Solheim B, Spjelkavik S, Olsen RA (2005) Nitrogen fixation in the high Arctic: role of vegetation and environmental conditions. Arct Antarct Alp Res 37:372–378